Data Sheet • GCI-AAV-293Ta packaging cell line • Catalog No. AA316

Description

GCI-AAV-293Ta packaging cells for AAV production using optimized packaging plasmids with AAV vector-based constructs.

Contents: 1 ml of GCI-AAV-293Ta packaging cells (2x10^{6}cells/ml). Low passage and authenticated.
Cell contains: Adenovirus E1 and SV-40 viral DNA sequences
Growth properties: Adherent
Organism: Homo sapiens (human)
Source organ: Kidney
Morphology: Epithelial

Growth medium: DMEM supplemented with 10% fetal bovine serum and Penicillin/Streptomycin
Temperature: 37.0°C

Thawing and Plating GCI-AAV-293Ta packaging cells

1. Remove the vial from liquid nitrogen freezer and immediately place it into a 37°C water bath. Remove the vial before the last bit of ice disappears to avoid overheating the cells. Wipe the vial with 70% ethanol.
2. Transfer thawed cells to a sterile 15ml or 50ml tube containing ~10 ml of complete growth medium (either cold or room temperature). Gently swirl the tube to mix.
3. Spin the cell suspension for 5 minutes at 300xg.
4. Carefully discard supernatant as completely as possible.
5. Tap the tube gently to dislodge and loosen the cell pellet.
6. Add ~10 ml of complete growth medium, and swirl the tube to suspend the cells.
7. Transfer the cell suspension to a 10cm culture dish or other appropriate culture vessel. The culture dish or vessel must be plasma tissue-culture treated.

   Important Note: The GCI-AAV-293Ta packaging cells were frozen in a DMSO containing medium. DMSO is toxic to cells and must be washed off before seeding the cells into a culture vessel. If DMSO is not removed, the cells will not attach and massive cell death will follow. If the culture dish is not plasma tissue-culture treated, the cells will not attach either.

Passaging

1. Discard culture medium.
2. Briefly rinse the cell layer with PBS.
3. Add 0.05% trypsin-EDTA solution to dish (just enough to cover the bottom of dish) and observe cells under an inverted microscope until the cell layer is dispersed.
4. Add >3× volume of complete growth medium (For example, add 9 ml or more of complete growth medium if 3 ml of trypsin-EDTA solution is used in step 3). Aspirate cells by gentle pipetting.
5. Transfer appropriate aliquots of the cell suspension to new culture vessels.
6. Incubate cells at 37°C with 5% CO_2.

Subcultivation ratio: Subcultivation ratio of 1:4 to 1:8 is recommended
Medium renewal: Every 2 to 3 days
Freeze medium: Complete growth medium supplemented with 8% (v/v) DMSO
Storage temperature: Liquid nitrogen vapor phase

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