



## NileHiFi® Long Amplicon PCR Kit

Cat. No. PC002 (50 reactions)

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## I. Introduction

NileHiFi® Long Amplicon PCR Kit is capable of amplifying templates up to 18kb of human gDNA, 30kb of  $\lambda$  phage DNA and other long genomic DNA. This kit is also recommended for amplifying GC rich or complex targets. NileHiFi® Long Amplicon PCR Kit is the ideal choice for generating long PCR products with high sensitivity and processivity.

## II. Contents and Storage

Catalog#	Component	Size	Storage
PC002-01	Long Amplicon DNA Polymerase (1U/ $\mu$ l)	100 $\mu$ l	-20°C
PC002-02	2× Long Amplicon PCR Mix	1.3 ml	
PC002-03	50mM Mg <sup>2+</sup>	50 $\mu$ l	

Avoid repeated freezing-thawing.

### DNA Polymerase Unit Definition

One unit is defined as the amount of enzyme that will incorporate 10 nmol of dNTP into acid-insoluble material in 30 minutes at 75°C.

## III. Important Notes:

1. Mix reagents thoroughly by gently inverting tubes several times while avoiding bubbles, and then briefly centrifuge before use.
2. Store the kit at -20°C. Avoid storage or leaving reagents at 4°C or room temperature. Avoid light exposure at all times.
3. Avoid repeated freezing-thawing.
4. Follow standard procedures for PCR to avoid nucleic acid contamination and non-specific amplification.
5. Read all procedures before setting up the PCR reaction.

## IV. Procedure

### 1. Primer design

- DNA fragment  $\leq$ 8 kb

Use professional primer design software or web design primers such as

PrimerPremier 5 or NCBI Primer Blast. If the DNA fragment is  $\leq$ 8kb, the length of the primer is recommended to be 20-25 bp, and the T<sub>m</sub> value is recommended to be higher than 55 °C. The T<sub>m</sub> value of the upstream and downstream primers should

avoid the difference, and the estimation should be made. The 3' end of the primer should avoid more than 3 binary repeats. sequence.

(2) DNA fragment  $\geq 8\text{kb}$

The primer length is recommended to be 25-30 bp, and the  $T_m$  value is recommended to be higher than 60 °C. The  $T_m$  value of the upstream and downstream primers should not be too high, and the GC content of the primer 3' should not be too high. Due to the amplification of macromolecular DNA fragments, it is recommended to optimize the primer design and use it preferentially.

## 2. Sample preparation (25/40/50 $\mu\text{l}$ )

	25 $\mu\text{l}$	40 $\mu\text{l}$	50 $\mu\text{l}$	Concentration
2 $\times$ Long Amplicon PCR Mix ( $\text{Mg}^{2+}$ 、dNTP plus) <sup>a</sup>	12.5 $\mu\text{l}$	20 $\mu\text{l}$	25 $\mu\text{l}$	1 $\times$
Forward Primer	0.4 $\mu\text{M}$	0.4 $\mu\text{M}$	0.4 $\mu\text{M}$	0.4 $\mu\text{M}$
Reverse Primer	0.4 $\mu\text{M}$	0.4 $\mu\text{M}$	0.4 $\mu\text{M}$	0.4 $\mu\text{M}$
Template	100 ng/100 pg	100 ng/100 pg	100 ng/100 pg	100 ng/100 pg
Long Amplicon DNA Polymerase(1U/ $\mu\text{l}$ )	1 $\mu\text{l}$	1.5 $\mu\text{l}$	2 $\mu\text{l}$	Various
ddH <sub>2</sub> O	Up to 25 $\mu\text{l}$	Up to 40 $\mu\text{l}$	Up to 50 $\mu\text{l}$	---

a. The optimal  $\text{Mg}^{2+}$  concentration of Long Amplicon DNA Polymerase is 1.5~3 mM. If the specific target band is not obtained, please adjust and optimize according to the FAQ of the section V.

b. If the template is gDNA, 50 ng or more should be used. If the template is plasmid DNA, 100 pg can be used.

c. Adjust the amount of polymerase added to different volumes of PCR reactions, such as 1  $\mu\text{l}$ /25  $\mu\text{l}$ ; 1.5  $\mu\text{l}$ /40  $\mu\text{l}$ ; 2  $\mu\text{l}$ /50  $\mu\text{l}$ .

## 3. PCR procedure

(1) DNA fragment  $\leq 8\text{kb}$ :

You can try a 3 step PCR first. If the specificity is not high enough, you can refer to the section VI for FAQ and solutions to make appropriate adjustments.

95°C <sup>a</sup>	2 min	
95°C	30 sec	} 30 cycles
55~60°C <sup>b</sup>	30 sec	
72°C	40 sec/kb	
72°C	5 min	
Keep in 4-10°C		

(2) For high GC% template or fragments larger than 8 kb:

it is recommended to use a 2 step PCR.

95°C	2 min	
95°C	30 sec	} 30 cycles
68°C <sup>c</sup>	40-50 sec/kb	
72°C	5 min	
Keep in 4-10°C		

a. Pre-denaturation time should be at least 2 minutes to completely melt DNA and activate the polymerase completely.

b. Stepwise temperature should be adjusted according to the T<sub>m</sub> value of the primers. If the T<sub>m</sub> value of the primers exceeds 55 °C, the continuous temperature can be set to 60 °C. If the T<sub>m</sub> value of the primers is lower than 55 °C, the continuous temperature can be set to 55 °C;

c. According to the T<sub>m</sub> value of the long fragment primer, the primer design principle is as described above, and the extension speed is as follows:

PCRproduct	≤12 kb	≥12kb	λ DNA
Extension speed	40 sec/kb	50 sec/kb	30 sec/kb

#### 4. Agarose gel electrophoresis

(1) PCR products should be detected by electrophoresis with 1×TAE buffer. Long-length PCR products are recommended to use low-concentration agarose gels and use low-voltage extension electrophoresis time to distinguish long fragments, for example:

Agarose : 0.6%

Electrophoresis condition: 90 V · 10 min ➡ 20 V · 6~16 h :

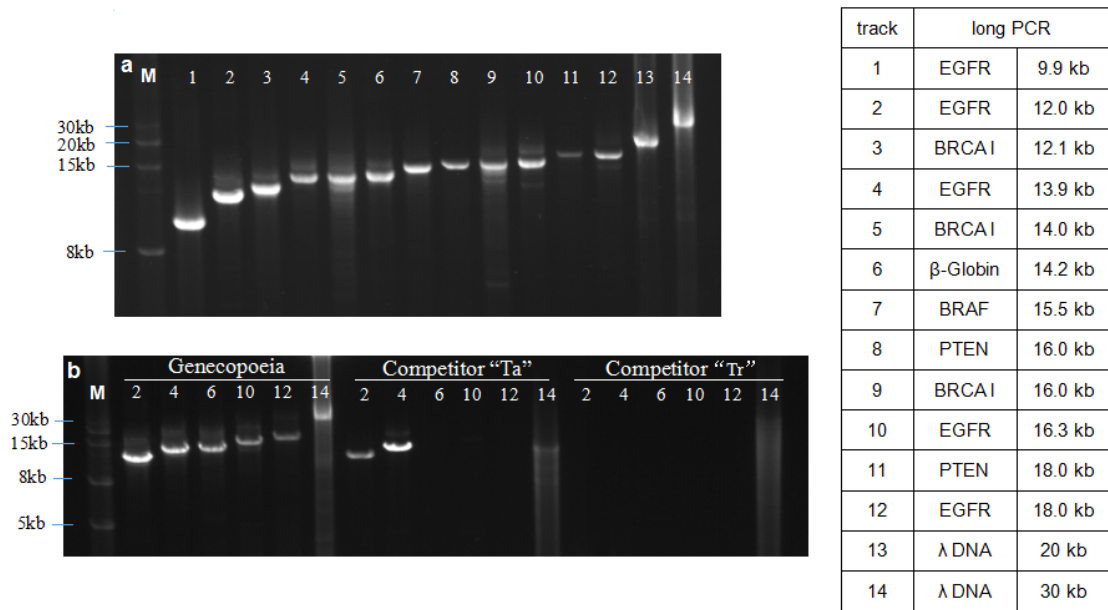


Figure 1. The NileHiFi® long amplicon PCR kit was used to amplify different long fragments of DNA. a. The length of the fragments is randomly selected and contains corresponding mutation targets (such as V600E, L858R, dE746-A750, etc.). b. Performance of GeneCopoeia's long amplicon PCR kit compared with equivalent kits from competitor "Ta" and competitor "Tr".

(2) Short-segment PCR products are recommended for using a 1% agarose gel, for example:

Agarose : 1%

Electrophoresis condition: : 140V · 20 min

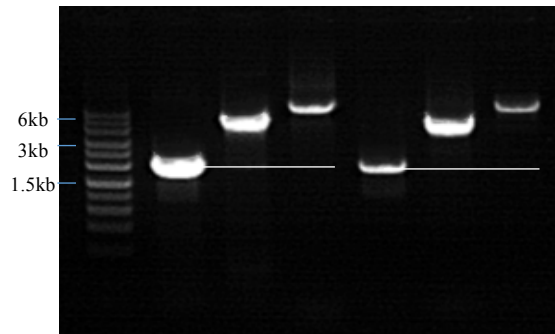


Figure 2. Genecopoeia's Long Amplicon PCR Kit short fragment amplification. Fragments are randomly selected and contain corresponding mutation targets (eg, V600E, dE746-A750, etc.).

## V. FAQ

Questions	Causes	Solutions
No specific fragments	Primer	Refer to the section IV Operation Flow 1. Primer design are optimized. It is recommended to design 2 or more primers for optimal use.
	Template	To increase the purity of the template, use appropriate

		amount of template DNA, the genomic DNA should not be less than 50 ng, and the plasmid DNA should be 100 pg.																
	Concentration of Mg <sup>2+</sup>	It is recommended to optimize the reaction system by appropriately increasing the final concentration of Mg <sup>2+</sup> by a gradient of 0.5 mM.																
	PCR Program	<p>1. If the 3-step PCR result is not satisfactory, try 2-step PCR; Slowdown PCR increases specificity</p> <table style="width: 100%; border-collapse: collapse;"> <tr> <td style="text-align: center;">95°C</td> <td style="text-align: center;">2 min</td> <td rowspan="3" style="font-size: 2em; vertical-align: middle;">}</td> <td rowspan="3" style="vertical-align: middle;">15 cycles</td> </tr> <tr> <td style="text-align: center;">95°C</td> <td style="text-align: center;">30 sec</td> </tr> <tr> <td style="text-align: center;">x=70°C · x=x-1</td> <td style="text-align: center;">20 sec</td> </tr> <tr> <td style="text-align: center;">72°C</td> <td style="text-align: center;">40-50 sec/kb</td> <td rowspan="3" style="font-size: 2em; vertical-align: middle;">}</td> <td rowspan="3" style="vertical-align: middle;">20 cycles</td> </tr> <tr> <td style="text-align: center;">95°C</td> <td style="text-align: center;">30 sec</td> </tr> <tr> <td style="text-align: center;">68°C</td> <td style="text-align: center;">40-50 sec/kb</td> </tr> </table>	95°C	2 min	}	15 cycles	95°C	30 sec	x=70°C · x=x-1	20 sec	72°C	40-50 sec/kb	}	20 cycles	95°C	30 sec	68°C	40-50 sec/kb
95°C	2 min	}	15 cycles															
95°C	30 sec																	
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72°C	40-50 sec/kb	}	20 cycles															
95°C	30 sec																	
68°C	40-50 sec/kb																	
Smear	Primer T <sub>m</sub> value	Refer to the section IV Operation Flow 1. Primer design principles are optimized. It is recommended to design 2 or more primers for optimal use.																
	Polymerase	Adjust the amount of polymerase, such as 0.6 μl / 25 μl or 1.5 μl/25 μl °.																
	Primer concentration	Adjust from 0.2 μM to 0.4 μM.																
	Extension time	Properly reduce the extension time.																
	PCR program	Reduce the number of cycles appropriately, for example 25~30.																
Low specific product yield	Template	Increase the amount of template appropriately, for example 200 ng																
	Extension time	Extend the time (1 min/kb) if the conditions are good specificity and no serious smear phenomenon.																
	PCR program	The number of cycles can be set to 32~35, which can increase the yield of PCR products.																

## **Limited Use License**

Following terms and conditions apply to use of NileHiFi® Long Amplicon PCR Kit. If the terms and conditions are not acceptable, the Product in its entirety must be returned to GeneCopoeia within 5 calendar days. A limited End-User license is granted to the purchaser of the Product. The product shall be used by the purchaser for internal research purposes only. The Product is expressly not designed, intended, or warranted for use in humans or for therapeutic or diagnostic use. The Product must not be resold, repackaged or modified for resale, or used to manufacture commercial products without prior written consent from GeneCopoeia. This Product should be used in accordance with the NIH guidelines developed for recombinant DNA and genetic research. Use of any part of the Product constitutes acceptance of the above terms.

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