

GeneHero[™] Cas9 Nuclease

Catalog No.	GE001	Cas9-NLS	25 µg
-	GE002	Cas9-NLS	100 µg
	GE003	Cas9-NLS-His	25 µg
	GE004	Cas9-NLS-His	100 µg

User Manual

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I. Description

GeneHero[™] Cas9 nuclease is the recombinant *Streptococcus pyogenes* Cas9 (wt) protein, purified from *E. coli*. Cas9 protein forms a ribonucleoprotein (RNP) complex with guide RNA *in vitro* that catalyzes site-specific double strand breaks that when repaired by non-homologous end joining creates small insertions and deletions. Incorporation of nuclear localization signals (NLS) at the N-terminal enhances the rate of genome editing by facilitating its delivery to the nucleus. Cas9 RNPs can cleave genomic targets with similar or higher efficiency as compared to Cas9/sgRNA plasmids. RNPs can be delivered to the cell as functional complexes and do not need transcription and translation; it is cleared rapidly from the cell that may increase CRISPR specificity and reduce off-target mutations.

II. Contents and Storage

Contents and storage recommendations for the GeneHero[™] Cas9 Nuclease are provided in the following table.

Catalog no.	Content	Conc.	Amount	Storage	
GE001	Cas9 protein with NLS sequence		25 μl (25 μg)		
GE002	Cas9 protein with NLS sequence	1	100 µl (100 µg)	-20 ºC	
GE003	Cas9 protein with NLS sequence and C-terminal 6X His-tag	- 1 μg/μl	25 μl (25 μg)	-20 ≌C	
GE004	Cas9 protein with NLS sequence and C-terminal 6X His-tag		100 µl (100 µg)		

Storage/dilution buffer: 10 mM Tris-HCl , 0.3 M NaCl , 1 mM DTT , 0.1 mM EDTA, 50 % Glycerol, pH =7.4

Store the GeneHero[™] Cas9 Nuclease at -80 ^QC until required for use. It is recommend to aliquot the Cas9 Nuclease as needed upon receiving the products.

Maintain RNase-free conditions by using RNase-free reagents, tubes, and barrier pipette tips while setting up your experiments.

Protocol: Cas9 protein cleavage of target DNA fragments in vitro

Overview

The protocol describes the *in vitro* digestion of DNA fragments using the Cas9/sgRNA RNP complex. This method can be used to validate the activity of sgRNA before application in *in vivo* studies.

Materials required but not provided

Synthetic sgRNA or *in vitro* transcribed (IVT) sgRNA DNA substrate containing the target sequence (plasmids, PCR products, or synthetic DNA oligo duplex) Cas9 protein storage/dilution buffer: 10 mM Tris-HCl , 0. 3 M NaCl , 1 mM DTT , 0.1 mM EDTA , 50 % Glycerol, (pH 7.4) 10X Reaction Buffer: 1 M NaCl, 500 mM Tris-HCl, 100 mM MgCl2, 1mg/ml BSA, pH 7.9 at room temperature. Proteinase K (20 mg/ml) Nuclease-free water

Tips

- Ensure the DNA substrate contains the 20 nt sgRNA targeting sequence, followed by the Cas9
 PAM site (NGG);
- Prepare DNA substrate with the target sequence by diluting the stock with nuclease-free water on ice.
 - Note: for synthetic DNA oligo duplex (≤100 bp), prepare 20-50 μM DNA substrate; for PCR fragments (100-2000 bp), prepare 20-500 nM DNA substrate; for linearized plasmid (≥ 2000 bp), prepare 10-20 nM DNA substrate.

The final concentration of DNA substrate in the reaction can range from 2 nM to 5 μ M depending on its format (linearized plasmid, PCR fragment) and length;

 The molar ratio of Cas9: sgRNA: DNA substrate should be at least 5: 5: 1 to obtain optimal cleavage efficiency;

Procedure

- Prepare $1 \mu M$ sgRNA by diluting the stock with nuclease-free water on ice.
- Dilute Cas9 nuclease to 160 ng/ μ l (about 1 μ M) using dilution buffer.
- Assemble the reaction at room temperature in the following order:

Component	Volume	Final Concentration	
Nuclease-free water	22 µl		
10X Reaction buffer	3 µl		
1μM sgRNA	1 µl	about 30 nM	
GeneHero™ Cas9 Nuclease	1 μl	about 30 nM	
(160 ng/μl)	τµι		
Reaction Volume	27 μl		
Pipette to mix and incubate for 10 min at 25 °C			
60 nM DNA substrate	3 µl	6 nM	
Total reaction volume	30 µl		

Mix thoroughly and incubate at 37 °C for 15~30 min.

- Add 1 μl of Proteinase K to the reaction, mix thoroughly
- Incubate at room temperature for 10 min, or at 56^oC for 10 min to achieve better effect.
- Analyze the digestion by agarose gel electrophoresis or Fragment Analyzer.

Protocol: Cas9 protein for genome modification in mammalian cell lines

Overview

This protocol describes the genome modification of mammalian cell line by lipid-based transfection of the Cas9/sgRNA RNP complex. Validation of the mutations can be conducted with the T7E1 assay.

Materials required but not provided

Synthetic sgRNA or *in vitro* transcribed (IVT) sgRNA CRISPR-Fectin[™] Transfection Reagent (GeneCopoeia, Cat# EF015) Opti-MEM[™] I Reduced Serum Medium (Life Technologies, Cat#31985-088) Nuclease-free water 6-, 24- or 96-well plate Rnase-free tips, tubes, etc.

Tips

- The optimal cell density for transfection varies for different cell lines based on cell size and growth characteristics. In general, a cell confluence of 30-50% on the day of transfection is recommended for lipid-based transfection.
- The molar ratio of Cas9: sgRNA should be at least 1: 1.3 to obtain optimal cleavage efficiency. It is
 recommended to optimize the dosage of RNP for transfection based on the cell line.

Procedure

Day 0. Seed cells

 If the cells are from a recent liquid nitrogen stock, passage the cells at least 2 times before transfection.

 The day before transfection, trypsinize and count the cells. Adjust the cell density and media volume according to the table below. Do not include antibiotics.

	6-well	24-well	96-well
Cell number per well	around 6 x 10 ⁵ cells	around 1 x 10 ⁵ cells	around 2.5 x 10 ⁴ cells
Volume of media per well	2 ml	0.5 ml	100 μl

Day 1.

 The number of cells plated in each well should be about 30%~50% confluence on the day of transfection.

Cas9-sgRNA RNP preparation

- 1. Thaw Cas9 protein with NLS sequence and sgRNA on ice. Dilute Cas9 protein using suitable buffer as needed. Dilute sgRNA using nuclease-free water.
- 2. For each well, mix sgRNA, Cas9 Nuclease and Opti-MEM[™]I Reduced Serum Medium according to the table below. Mix well using pipette, reduce bubbles during pipetting.

	6-well	24-well	96-well
sgRNA	32.5 pmol	6.5 pmol	1.3 pmol
Cas9 Nuclease	4000 ng (25 pmol)	800 ng (5 pmol)	160 ng (1 pmol)
Opti-MEM™ I Medium	125 μl	25 μl	5 µl

3. Incubate at room temperature for 5 min to assemble the RNP complexes.

Transfect the RNP complex

4. Dilute CRISPR-Fectin[™] transfection reagent in Opti-MEM[™] I Medium according to the table below. Mix well.

	6-well	24-well	96-well
CRISPR-Fectin™	7.5 μl	1.5 μl	0.3 μl
Opti-MEM™ I Medium	125 μl	25 μl	5 µl

- 5. Incubate the CRISPR-Fectin Max[™] transfection reagent in Opti-MEM[™] I Medium at room temperature for 1 minute.
- 6. Add the diluted CRISPR-Fectin Max[™] transfection reagent to the Cas9-sgRNA RNP mixture. Mix well by pipetting.
- 7. Incubate the mixture of RNP and transfection reagent at room temperature for 15 to 20 min, do not exceed 30 min.
- 8. Add the mixture to the cells according to the table and mix gently by rocking the plate back and forth.

	6-well	24-well	96-well
RNP/CRISPR-Fectin™	250	50 JU	10 ul
mixture	250 μl	50 μl	10 µl

9. Incubate the cells at 37° C in a CO₂ incubator for 2-3 days until they are ready to be assayed.

 It is recommended to use the IndelCheck[™] CRISPR insertion or deletion detection system (GeneCopoeia, Cat# IC001, IC002) to check RNP transfection efficiency.

VI. Limited Use License and Warranty

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