

# Harnessing CRISPR For Activation of Gene Expression

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**Presenter:**

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GeneCopoeia, Inc.**

# GeneCopoeia Products and Services

## Functional Genomics & Cell Biology

<b><i>Clones</i></b>	<b><i>Viral systems</i></b>	<b><i>Stable Cell Lines</i></b>	<b><i>Kits &amp; reagents</i></b>	<b><i>Fluorescent detection</i></b>
ORF Promoter miRNA CRISPR shRNA	Lentivirus AAV	CRISPR-Cas9 stable cell lines Labeled cancer cell lines Cancer biomarker mutant cell lines	Transfection Luciferase FISH probes Indel detection Cloning	Cell function assays Nucleic acid detection Cell structure probes Fluorescent dyes



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# Outline

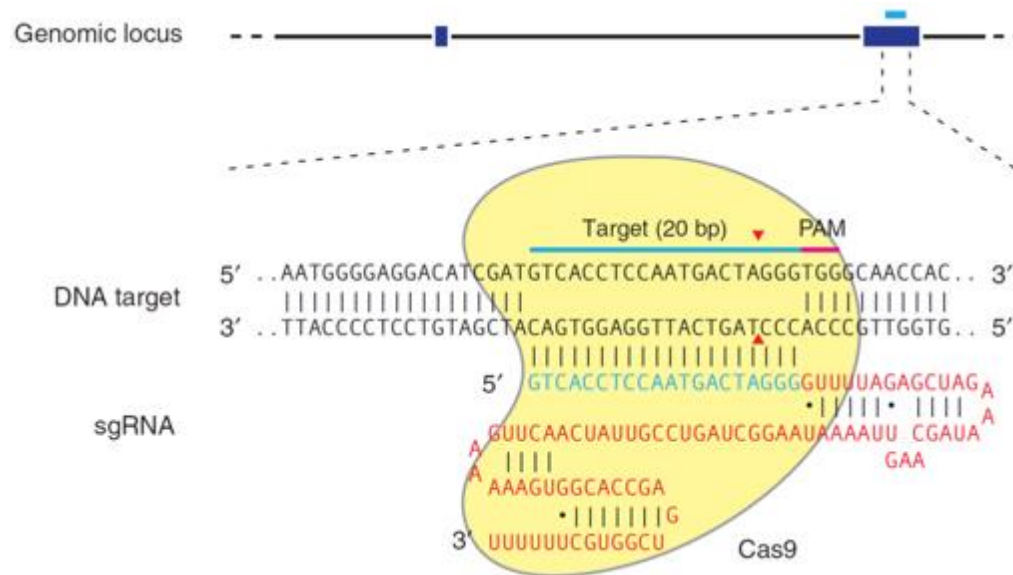
- ❖ CRISPR technologies & applications
- ❖ Introduction to CRISPR activation
- ❖ Delivery methods
- ❖ Preparation
- ❖ Screening

# Outline

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# CRISPR-Cas9 genome editing technology

## CRISPR-Cas9: RNA-guided endonuclease



- ❖ 17-20 nt single guide RNA (sgRNA) guides Cas9 nuclease to target site.
- ❖ Requires NGG “PAM” site immediately downstream of sgRNA target sequence.
- ❖ Cas9-sgRNA complex makes DSB 3 nt upstream of PAM.

Ran, et al. (2013). Nature Protocols 8, 2281

# CRISPR-Cas9 genome editing technology

## Technological variants

Technology	Description	PAM	Applications
SpCas9	Unmodified Cas9 from <i>Streptococcus pyogenes</i>	NGG	Gene knockout, knock-in, tagging, mutagenesis
SpCas9-NG	Mutagenized SpCas9 with relaxed (NG) PAM requirement	NG	Gene knockout, knock-in, tagging, mutagenesis
Cas9-HF, eSpCas9	Mutagenized, high-fidelity SpCas9 for reduced off-targeting	NGG	Gene knockout, knock-in, tagging, mutagenesis
HiFi Cas9	Mutagenized, high-fidelity SpCas9 for reduced off-targeting	NGG	Gene knockout, knock-in, tagging, mutagenesis
Cpf1 (Cas12a)	Nucleases from <i>Acidaminococcus</i> sp. BV3L6 or <i>Lachnospiraceae</i> bacterium ND2006. Smaller than SpCas9, no requirement for tracrRNA, staggered DSB, lower off-targeting.	T/CTTN (5')	Gene knockout, knock-in, tagging, mutagenesis
SaCas9	Cas9 from <i>Staphylococcus aureus</i> . Smaller than SpCas9, usually used with adenoassociated virus (AAV)	NNGRRT	Gene knockout, knock-in, tagging, mutagenesis
dCas9	Nuclease-dead Cas9 for tethering to effectors	NGG	Gene mutagenesis, chromatin modification, transcriptional activation/repression
Cas13	Nuclease from <i>Leptotrichia wadei</i> . Targets RNA. 20-28 nt spacer	None	RNA knockdown or mutagenesis

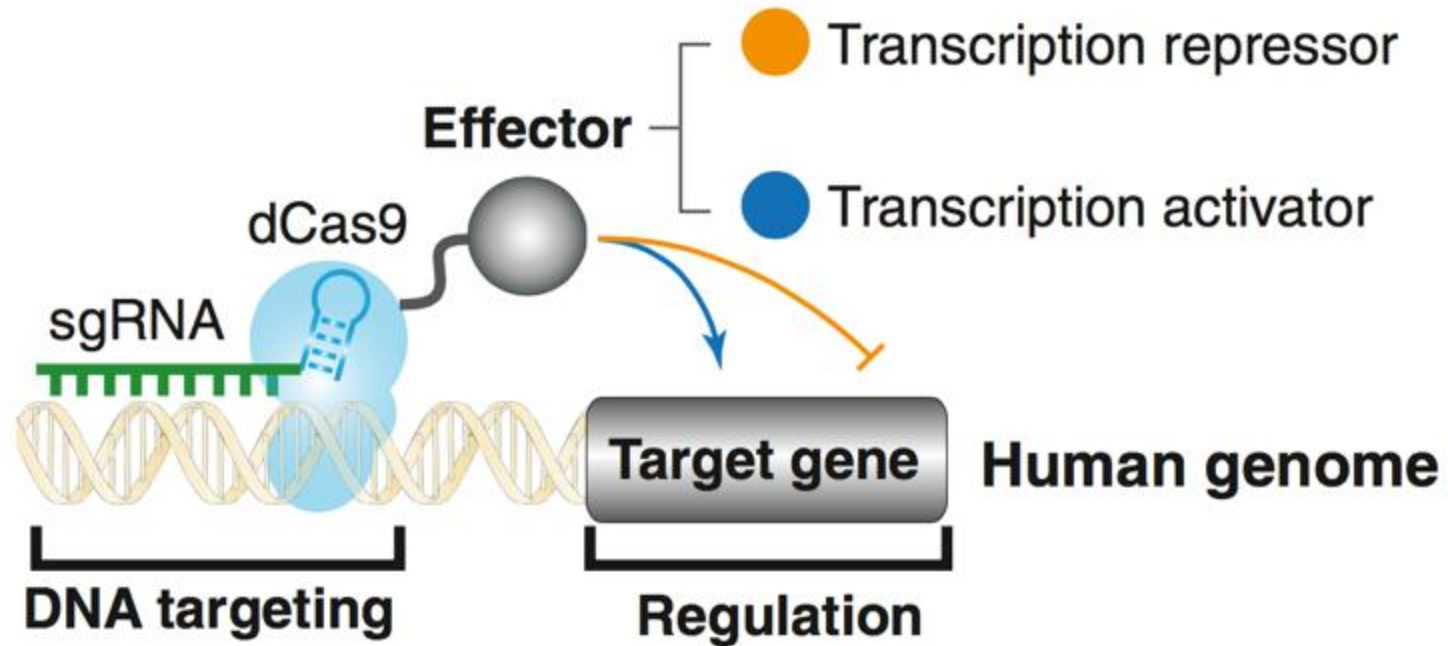


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- ❖ **Introduction to CRISPR activation**
- ❖ Delivery methods
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# CRISPR-Cas9 applications

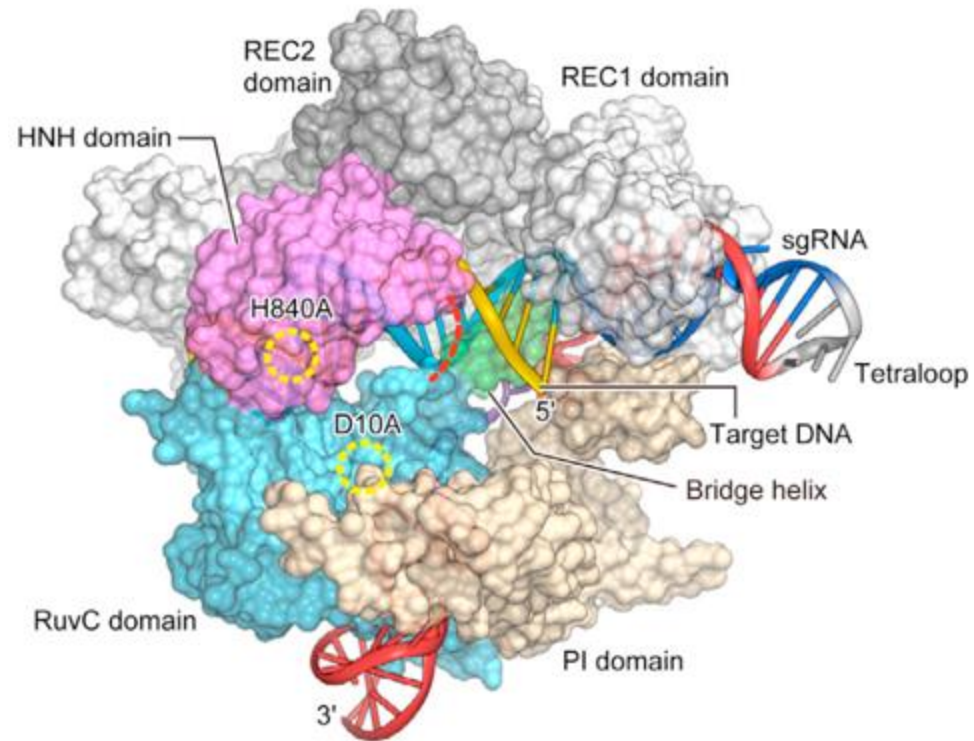
Gene activation or repression



Gilbert, et al. (2013). Cell 154, 442

# CRISPR-Cas9 applications

## Nuclease-dead Cas9



Nishimasu, et al. (2014). Cell 156, 935

# Cas9-mediated gene activation

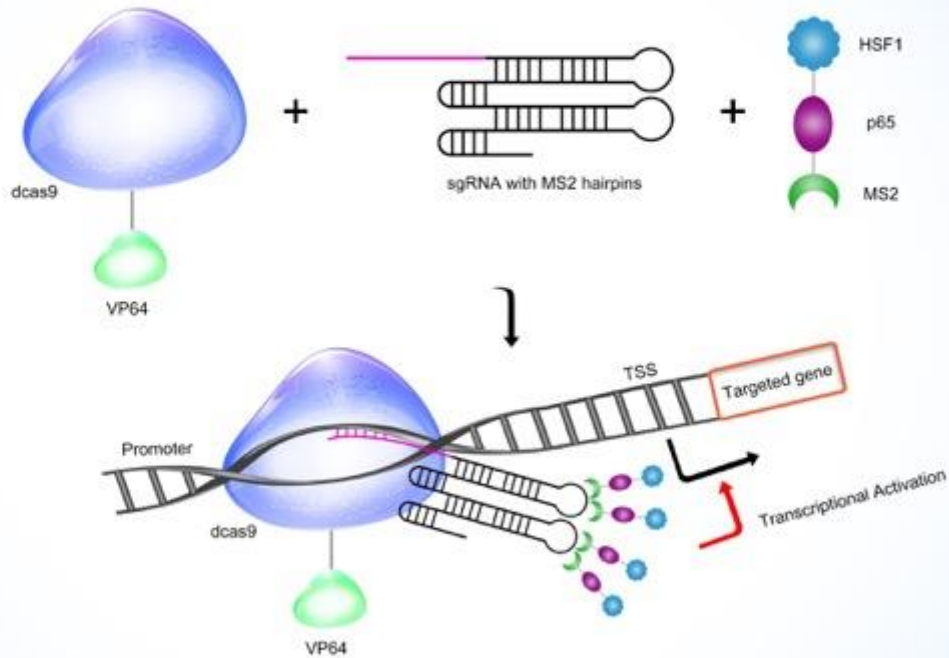
## Why use CRISPR for this?

- ❖ Can overexpress genes using open reading frame (ORF) clones, but...
- ❖ ORF clones use artificial promoters
- ❖ Not all protein isoforms are always covered
- ❖ Large ORFs sometimes too difficult to clone

# dCas9-SAM system

Konermann, *et al.*, (2015). *Nature* 517, 583

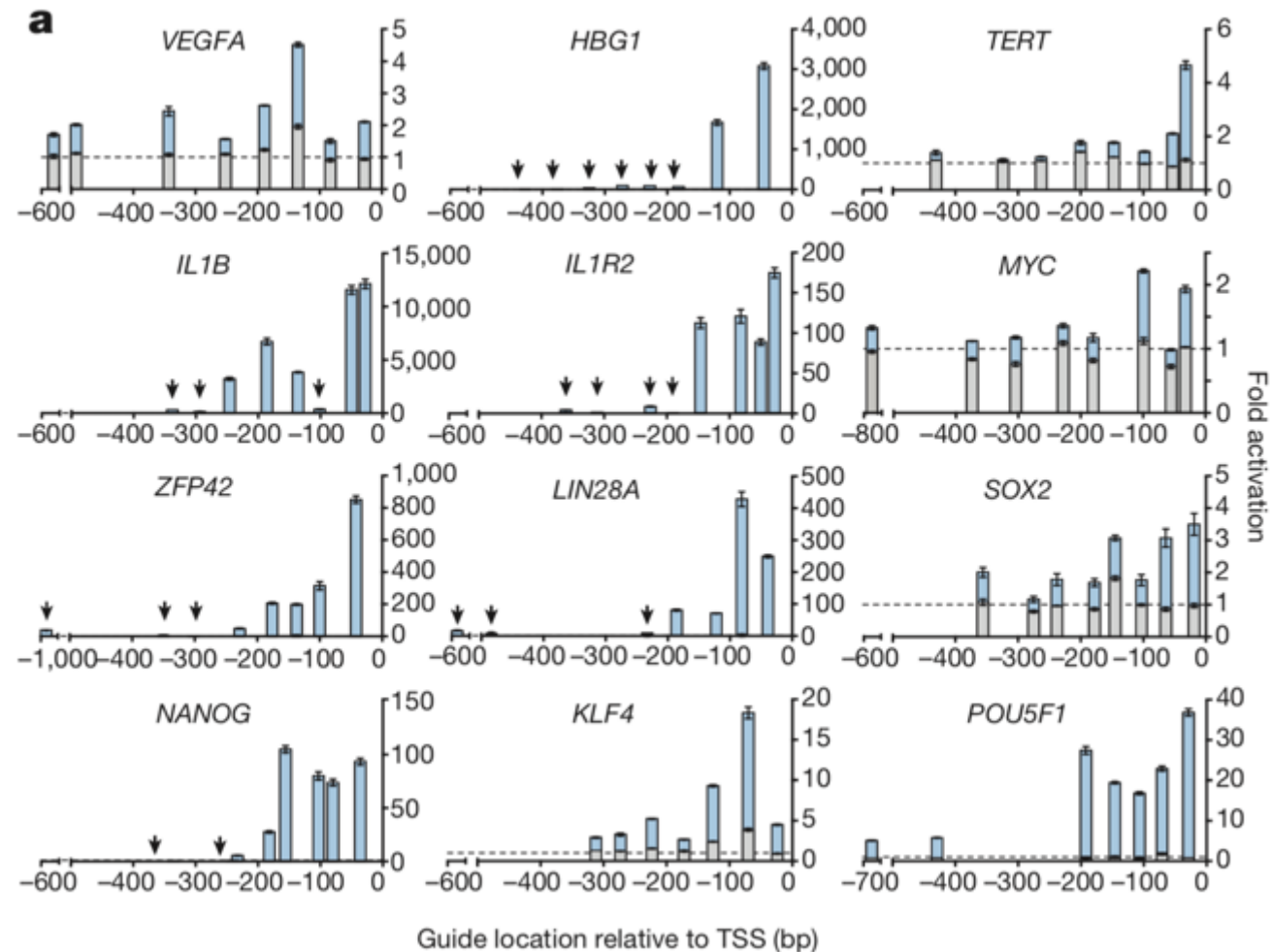
## **SAM Transcription Activation System**



- ❖ 3-component system: 1) dCas9-VP64 fusion; 2) HSF1-p65-MS2 fusion; 3) Hybrid sgRNA-apatamer
- ❖ Uses same type of 20 nt sgRNA sequence that recognizes NGG PAM as Cas9 nuclease for DSBs
- ❖ Optimal activation occurs with sgRNAs up to 200 bp 5' of transcription start site (TSS)

# dCas9-SAM system

Konermann, *et al.*, (2015). *Nature* 517, 583

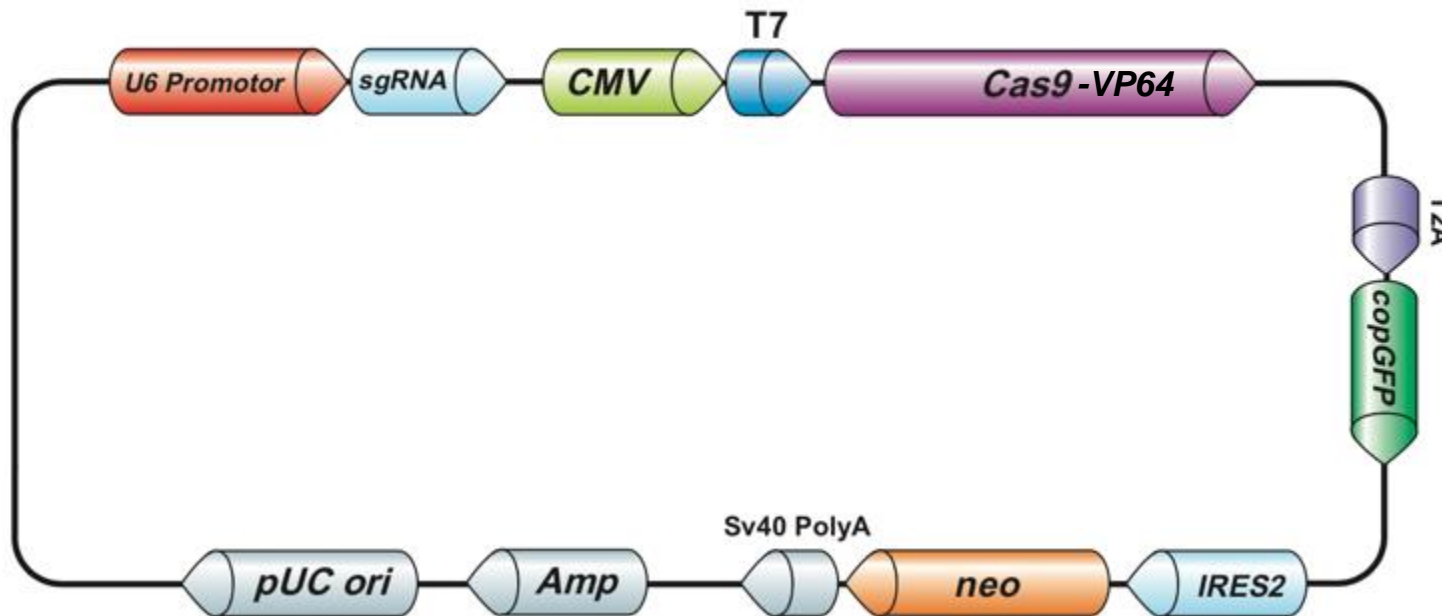


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- ❖ **Delivery methods**
- ❖ Preparation
- ❖ Screening

# CRISPR-Cas9 gene activation technology

## Delivery method 1: Plasmid DNA transfection



- ❖ GeneCopoeia Cas9 + sgRNA “All-in-one” plasmid
- ❖ Would also need separate plasmid expressing HSF1-p65-MS2 fusion
- ❖ Fluorescent reporter allows for sorting
- ❖ Selection marker can be used for stable integration
- ❖ Cas9 needs to be transcribed and translated

# CRISPR-Cas9 gene activation technology

## Delivery method 2: RNA transfection

- ❖ Co-transfect *in vitro* transcribed dCas9-VP64 mRNA, HSF1-p65-MS2 fusion, and sgRNA (aptamer modified)
- ❖ sgRNA can either be encoded on plasmid, *in vitro* transcribed, or synthetic
- ❖ Advantage: Plasmid-free, so cannot integrate
- ❖ dCas9-VP64 still needs to be translated

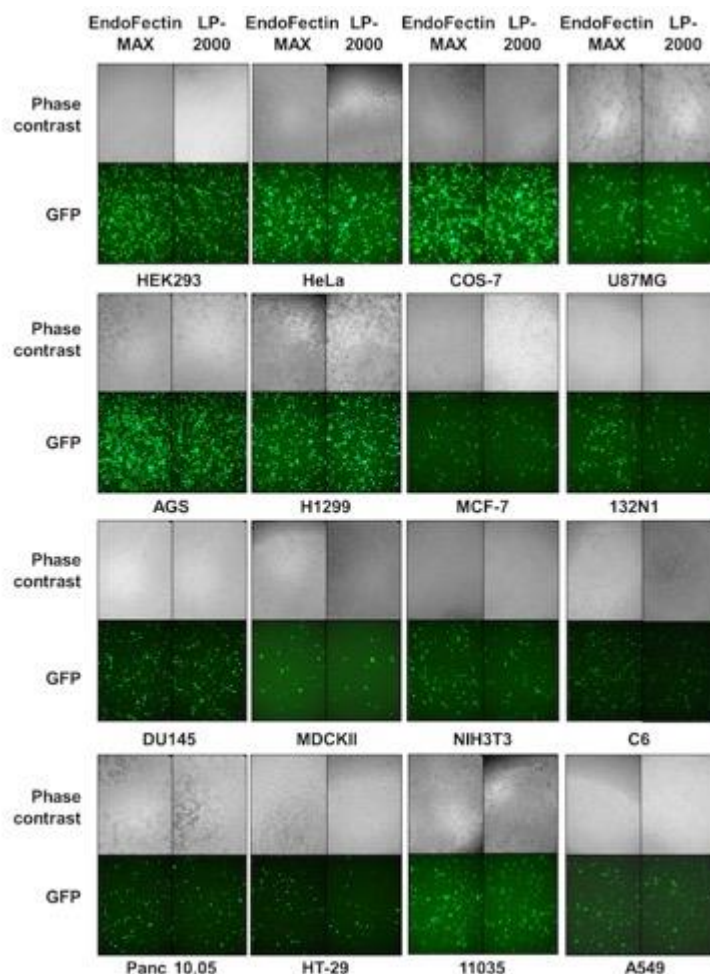
# CRISPR-Cas9 gene activation technology

## Delivery method 3: Protein transfection

- ❖ Pre-form complex of dCas9-VP64 protein and aptamer-modified sgRNA in vitro to form RNP
- ❖ Aptamer-modified sgRNA can be either chemically synthesized or *in vitro* transcribed
- ❖ Transfect cells with RNP complex using either lipid transfection reagent or electroporation.  
Co-transfect with HSF1-p65-MS2 mRNA
- ❖ Does not integrate, unlike plasmids

# CRISPR-Cas9 gene activation technology

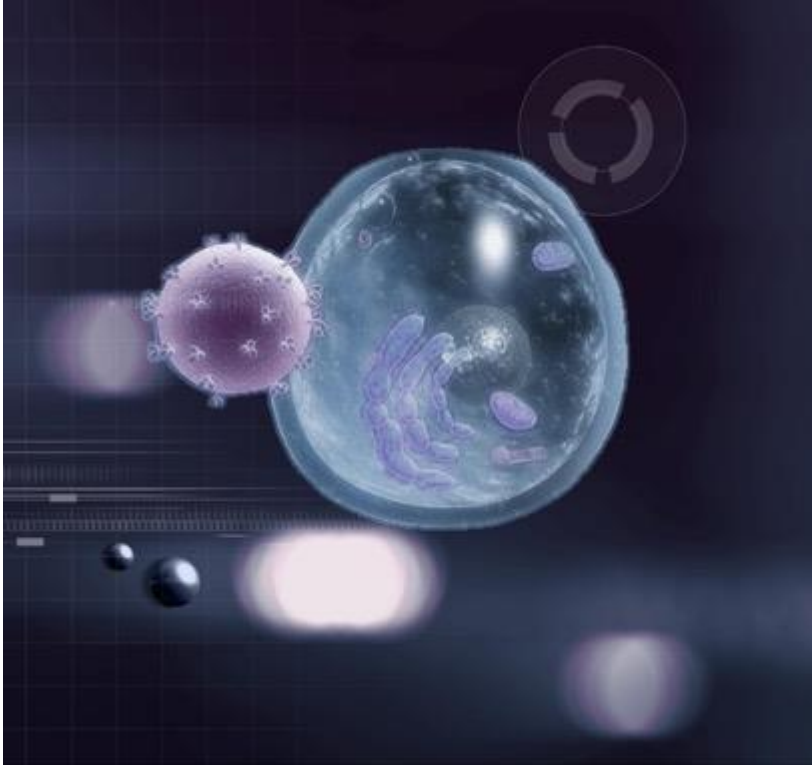
## EndoFectin™ Max: Robust transfection reagent



- ❖ Equal to or greater transfection efficiency compared with Lipofectamine™ 2000 or Lipofectamine™ 3000
- ❖ Especially robust with difficult-to-transfect cell lines
- ❖ Capable of efficient transfection of DNA, RNA, and protein

# CRISPR-Cas9 gene activation technology

## Delivery method 4: Lentivirus



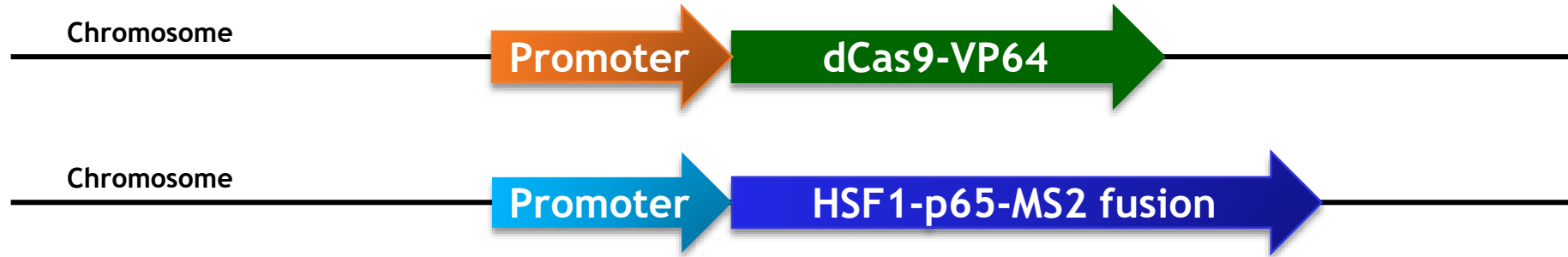
### Why lentivirus?

- ❖ DNA transfection not always possible or practical. Some cell lines difficult or impossible to transfect.
- ❖ Most mammalian cells support infection by engineered lentivirus



# CRISPR-Cas9 gene activation technology

## GeneHero™ Cas9a stable cell lines

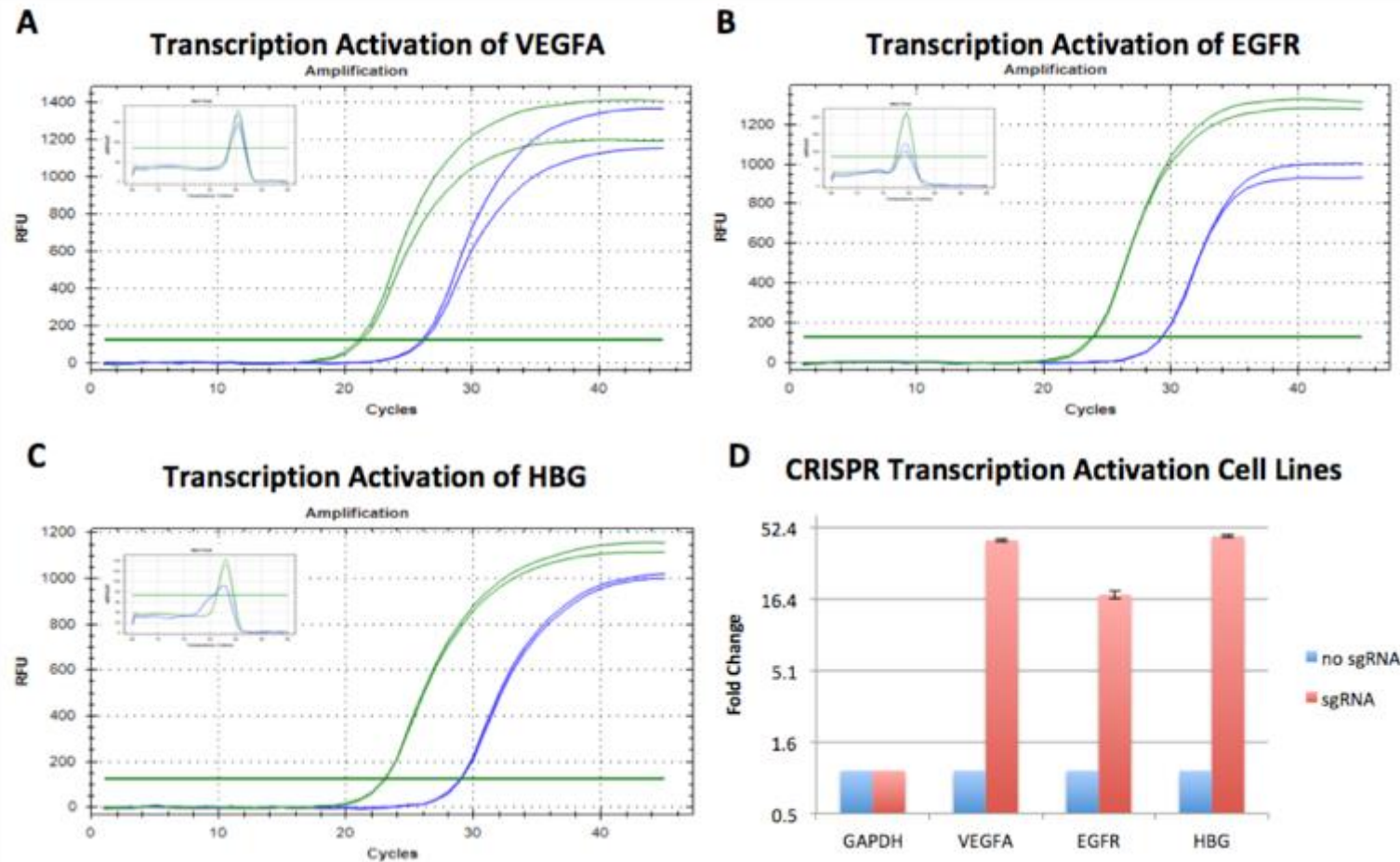


### Features

- ❖ Cell lines with dCas9-VP64 and HSF1-p65-MS2 stably integrated in the genome
- ❖ 2 pre-made cell lines available: HEK293 and MCF-7
- ❖ Functionally validated for CRISPR-mediated activation
- ❖ Ideal for lentiviral CRISPRa applications
- ❖ Need to provide sgRNA with aptamer

# GeneHero™ Cas9a stable cell lines

## SL302-HEK293

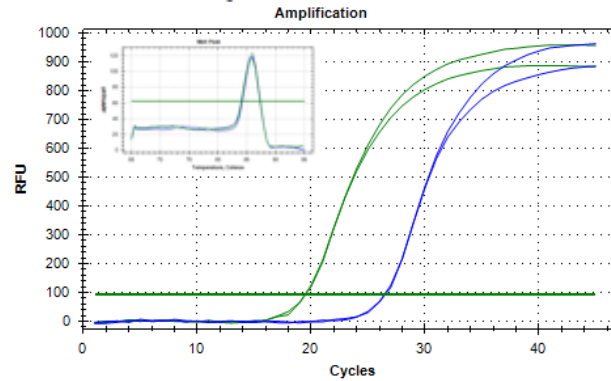


# GeneHero™ Cas9a stable cell lines

## SL504-MCF-7

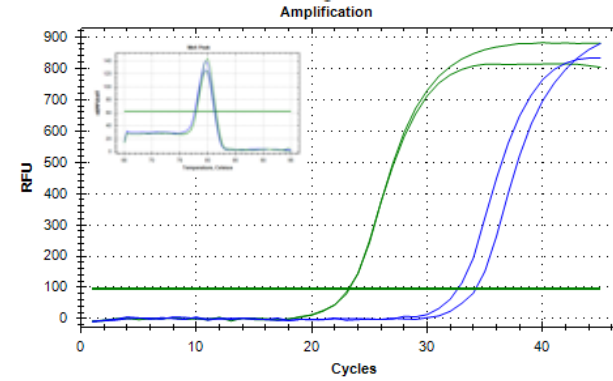
**A**

### Transcription Activation of VEGFA



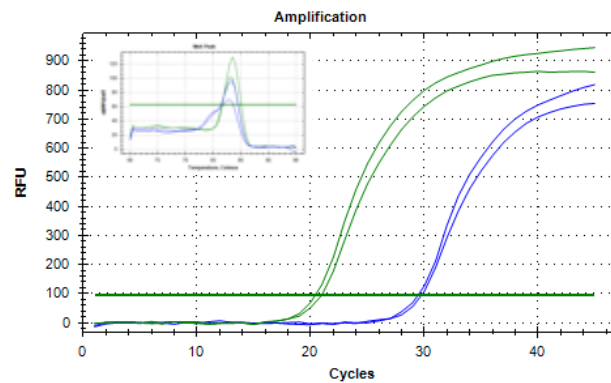
**B**

### Transcription Activation of EGFR



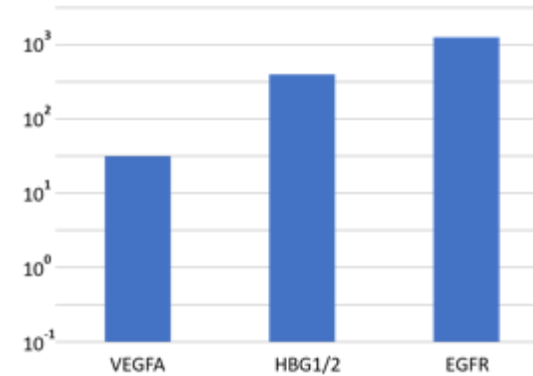
**C**

### Transcription Activation of HBG



**D**

### CRISPR Transcription Activation Cell Lines



# Outline

- ❖ CRISPR technologies & applications
- ❖ Strategy considerations
- ❖ Delivery methods
- ❖ **Preparation**
- ❖ Screening

# CRISPR-Cas9 gene activation technology

## Workflow: Preparation

Step 1. Determine the DNA sequence of the sgRNA target site in your cell line

- ❖ Necessary to ensure highest possible nucleotide match before sgRNA synthesis

# CRISPR-Cas9 gene activation technology

## Smart-Join™ Blunt-end PCR Cloning Kit

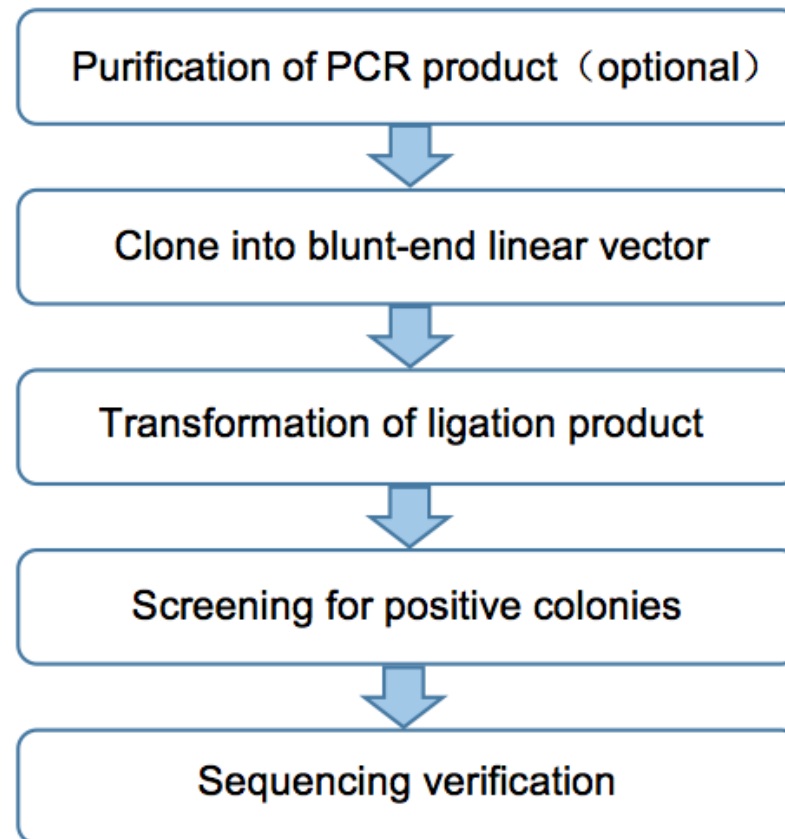
### Features

- ❖ Convenient method for high efficiency cloning of blunt-end PCR products
- ❖ Higher positive ligation rate and true positive colonies by the elimination of self-ligating products
- ❖ Cloning of blunt-end PCR products without additional sequences or restriction sites
- ❖ Ideal for sequencing regions of genomic DNA

# CRISPR-Cas9 gene activation technology

## Smart-Join™ Blunt-end PCR Cloning Kit

### Workflow



# CRISPR-Cas9 gene activation technology

## Workflow: Preparation

### Step 2: Validate sgRNA function in your cell line

- ❖ Not all sgRNAs are created equal. It is the best practice to try multiple target sites and pre-validate the best one
- ❖ Validate via qPCR

# CRISPR-Cas9 gene activation technology

## BlazeTaq™ SYBR green qPCR system

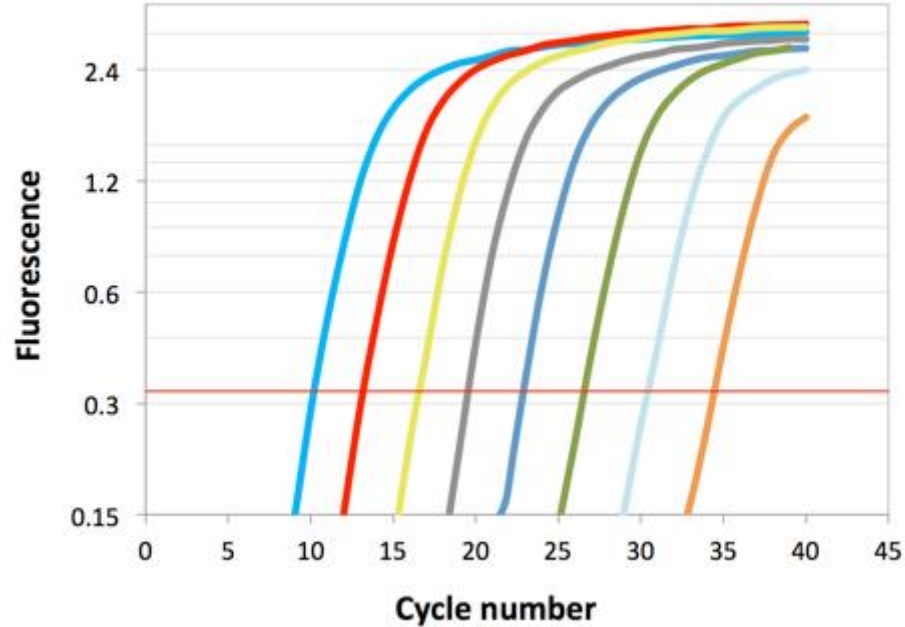
### Features

- ❖ Available as either 2-step (RT + qPCR separately) or 1-step (RT and qPCR in the same reaction) options
- ❖ Highly sensitive. Detects as little as 5 copies of DNA template or 0.1 pg of RNA template
- ❖ High specificity with minimal level of primer-dimer and non-specific product formation
- ❖ Antibody-modified polymerase provides faster heat activation than chemically modified enzyme (30 sec. vs. 15 min.)

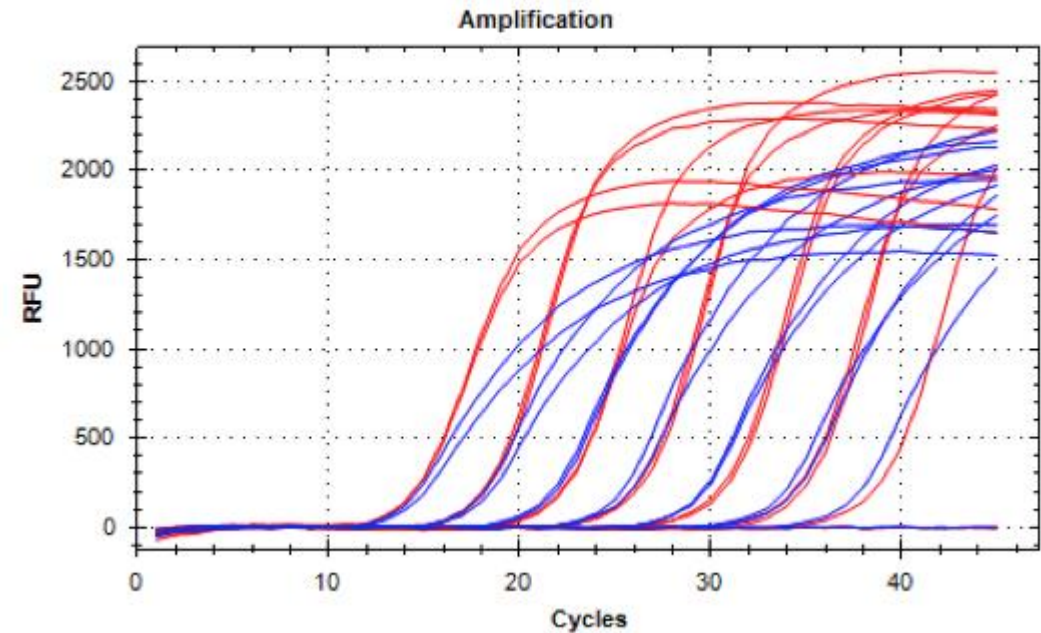
# CRISPR-Cas9 gene activation technology

BlazeTaq™ SYBR green qPCR system

Amplification Plot

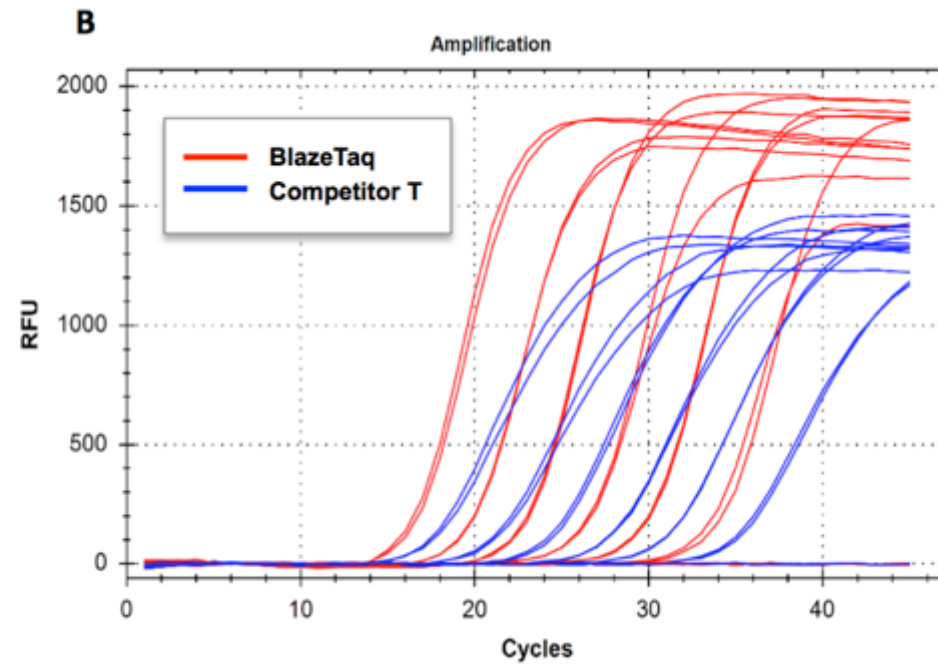
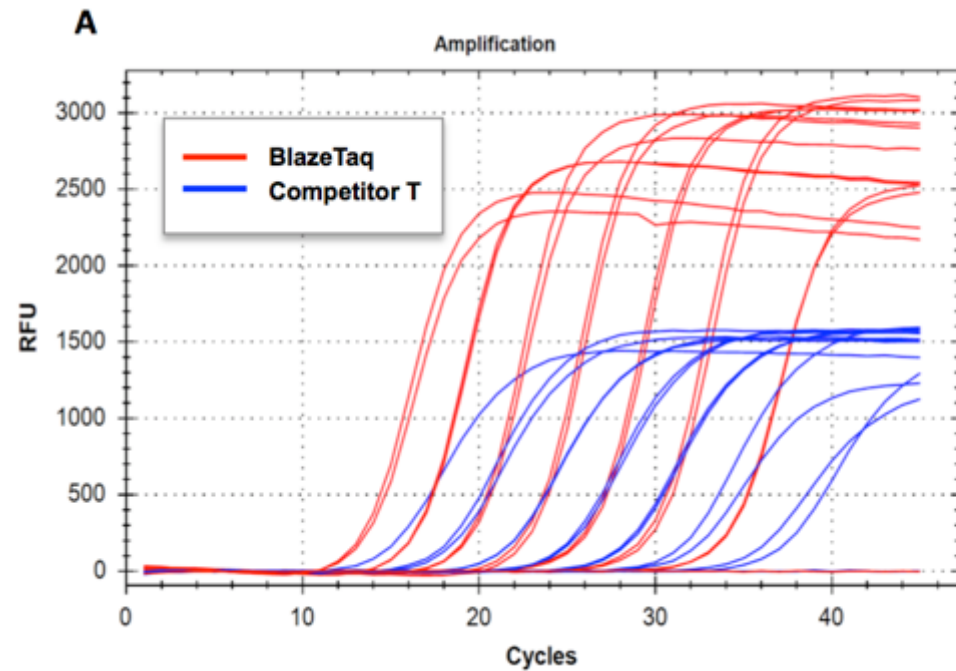


Performance Comparison with Competitor



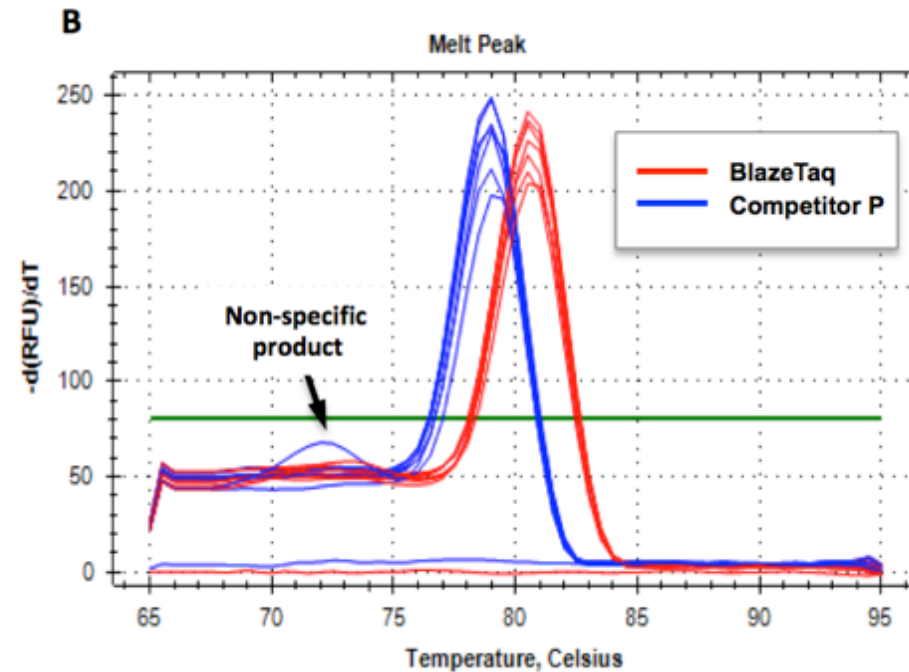
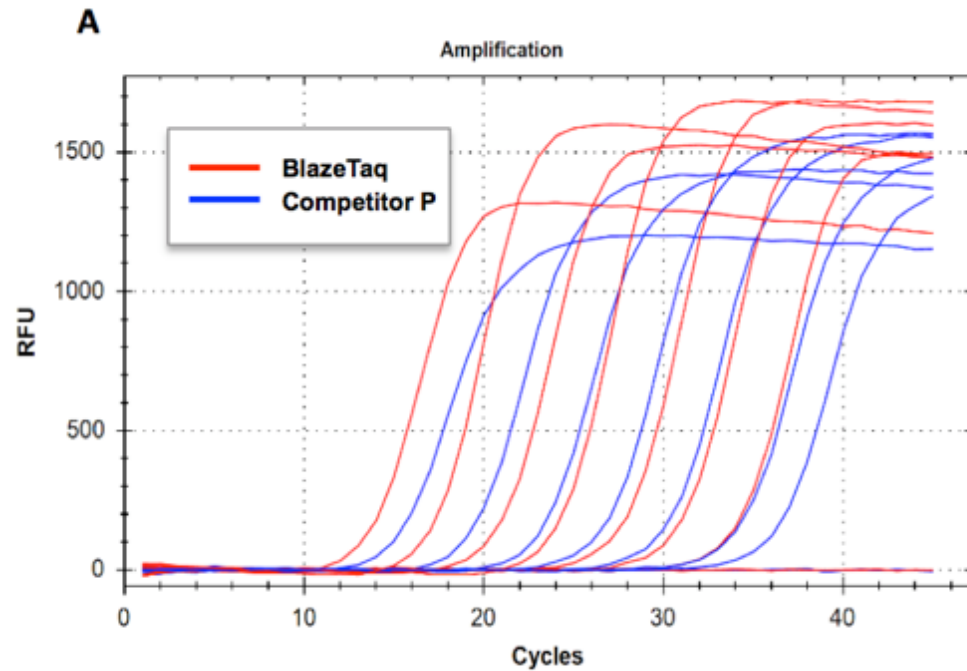
# CRISPR-Cas9 gene activation technology

BlazeTaq™ SYBR green one-step qPCR system



# CRISPR-Cas9 gene activation technology

BlazeTaq™ SYBR green one-step qPCR system

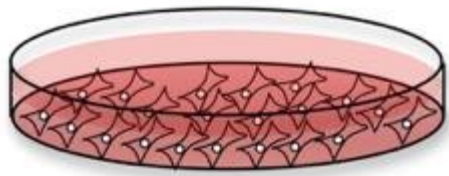


# Outline

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- ❖ **Screening**

# Screening

Isolate single clones post-transfection/transduction



**OR**



**OR**

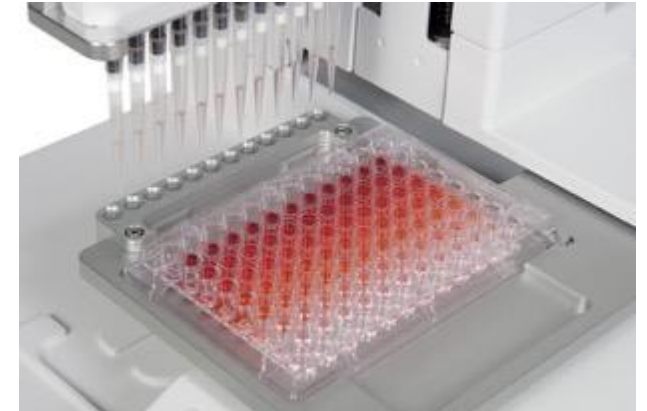


Plate for single colonies  
and pick off dish

Fluorescence sorting

Do serial dilutions in  
multi-well plates

- ❖ Minimizes potential effects of unwanted modifications resulting from random insertion, cell division or off-targeting

# Screening

Isolate single clones post-transfection/transduction

- ❖ Screen single clones using the BlazeTaq™ SYBR green qPCR kits to analyze for transcriptional activation

# Summary

- ❖ CRISPR-Cas9-mediated gene activation is a useful system for potentially overexpressing genes, and has distinct advantages over the use of ORF clones, especially for high-throughput screening
- ❖ Currently, the most robust CRISPR activation system in use is the dCas9 system of Konermann, et al.
- ❖ GeneCopoeia offers many products and services, including plasmid DNA clones, Cas9 activation cell lines, and robust qPCR reagents, to help you meet your CRISPR gene activation needs

# Thank You!

If you have any additional  
questions, please call

1-866-360-9531 x227

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Or visit us on the web:

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