

# Transferring an ORF from an EZShuttle<sup>™</sup> Gateway<sup>®</sup> PLUS ORF clone or ORFeome Shuttle Clone to a Destination Vector

## Reagents

EZShuttle<sup>™</sup> Gateway PLUS ORF clone or ORFeome Gateway entry clone 5X Reaction Buffer (GeneCopoeia Cat. No. RCBM-1001-020) EZRecombinase<sup>™</sup>LR mix (GeneCopoeia Cat. No. RCBM-1001-020) Linearized destination vector 50 mM TE buffer, pH 8.0 Proteinase K GCI-5α chemically competent *E. coli* cells (GeneCopoeia Cat. No. CC001) S.O.C. Medium LB plates with 100 µg/ml ampicillin

# Equipment

Autoclaved assorted pipet tips Pipetman pipets: P2, P20, P200, P1000 Autoclaved 1.5-ml microtubes 17 x 100 mm polypropylene tubes (Falcon 2059) 37°C and 42°C incubators

## Procedures

### 1. LR Reaction

Use the following procedure to perform an LR recombination reaction. For a positive control, use 100 ng (2 µl) of pENTR<sup>™</sup>-eGFP.

1.1 Add the following components to a 1.5-ml microcentrifuge tube at room temperature and mix:

Component	Volume
Entry clone (100–300 ng)	1–10 µl
Destination vector (150 ng/µl)	2 µl
5X Reaction Buffer	4 µl
TE buffer, pH 8.0	to 16 μΙ

- 1.2 Remove EZRecombinase<sup>™</sup> LR mix from −80°C and thaw on ice for about 2 minutes. Vortex briefly twice (2 seconds each time).
- 1.3 To each sample (Step 1.1 above), add 4 μl of EZRecombinase<sup>™</sup> LR mix to the reaction and mix well by vortexing briefly twice. Microcentrifuge briefly.
- 1.4 Return EZRecombinase<sup>TM</sup> LR mix to  $-80^{\circ}$ C storage immediately after use.
- 1.5 Incubate reactions at 25°C for 60 minutes.

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#### 2. Transformation

2.1 Transform 1 µl of each LR reaction into 50 µl of GCI-5α chemically competent *E. coli* cells. Incubate on ice for 30 minutes. Heat shock cells by incubating at 42°C for 30 seconds. Add 450 µl of S.O.C. Medium and incubate at 37°C for 1 hour with shaking. Plate 20 µl and 100 µl of each transformation onto selective plates.

**Note:** Any competent cells with a transformation efficiency of >1.0  $\times$  10<sup>8</sup> transformants/µg may be used.

2.2 Transform 1 μl of pUC19 DNA (10 ng/ml) or other suitable plasmid for use as a control into 50 μl of GCI-5α chemically competent *E. coli* cells as described above. Plate 20 μl and 100 μl on LB plates containing 100 μg/ml ampicillin

#### **Expected results**

An efficient LR recombination reaction will produce > 5000 colonies if the entire transformation is plated.

#### LB plates (per liter)

- 10.0 g Bacto-Tryptone
- 5.0 g Bacto-Yeast extract
- 5.0 g NaCl
- 15.0 g Agar

Adjust the pH to 7.0 with NaOH (~200 µl 5M NaOH). Autoclave (keep the top loosened to allow steam to vent) and cool to 50°C before adding antibiotics. Mix well and pour LB media into plates.

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