

## Data Sheet • 293Ta Lentiviral Packaging Cell Line • Catalog No. LT008

### Description

293Ta packaging cells for lentivirus production using optimized packaging plasmids with lentiviral vector-based constructs.

**Contents:** 1 ml of 293Ta lentiviral packaging cells ( $1.5 \times 10^6$  cells/ml). Low passage and authenticated.

**Cell contains:** Adeno and SV-40 viral DNA sequences

**Growth properties:** Adherent

**Organism:** Homo sapiens (human)

**Source organ:** Kidney

**Morphology:** Epithelial

**Growth medium:** DMEM supplemented with 10% fetal bovine serum and Penicillin/Streptomycin

**Temperature:** 37.0°C

### Thawing and Plating 293Ta Cells

1. Remove the vial from liquid nitrogen freezer and immediately place it into a 37°C water bath. Remove the vial before the last bit of ice disappears to avoid overheating the cells. Wipe the vial with 70% ethanol.
2. Transfer thawed cells to a sterile 15ml or 50ml tube containing ~10 ml of complete growth medium (either cold or room temperature). Gently swirl the tube to mix.
3. Spin the cell suspension for 5 minutes at 300xg.
4. Carefully discard supernatant as completely as possible.
5. Tap the tube gently to dislodge and loosen the cell pellet.
6. Add ~10 ml of complete growth medium, and swirl the tube to suspend the cells.
7. Transfer the cell suspension to a 10cm culture dish or other appropriate culture vessel. The culture dish or vessel must be plasma tissue-culture treated.

**Important Note:** The 293Ta packaging cells were frozen in a DMSO containing medium. DMSO is toxic to cells and must be washed off before seeding the cells into a culture vessel. If DMSO is not removed, the cells will not attach and massive cell death will follow. If the culture dish is not plasma tissue-culture treated, the cells will not attach either.

### Passaging

1. Discard culture medium.
2. Briefly rinse the cell layer with PBS.
3. Add 0.05% trypsin-EDTA solution to dish (just enough to cover the bottom of dish) and observe cells under an inverted microscope until the cell layer is dispersed.
4. Add >3x volume of complete growth medium (For example, add 9 ml or more of complete growth medium if 3 ml of trypsin-EDTA solution is used in step 3). Aspirate cells by gentle pipetting.
5. Transfer appropriate aliquots of the cell suspension to new culture vessels.
6. Incubate cells at 37°C with 5% CO<sub>2</sub>.

**Subcultivation ratio:** Subcultivation ratio of 1:4 to 1:8 is recommended

**Medium renewal:** Every 2 to 3 days

**Freeze medium:** Complete growth medium supplemented with 8% (v/v) DMSO

**Storage temperature:** Liquid nitrogen vapor phase

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