

VitroView™ Masson's Trichrome Stain Kit

Cat. No. VB-3016

Introduction

Masson's Trichrome Stain Kit is used to differentiate between collagen and smooth muscle in tumors, and the increase of collagen in diseases such as cirrhosis. It is a routine stain for liver and kidney biopsies. This kit can be used for FFPE and frozen tissue slides.

Kit Components

1. Bouin's solution -----250 ml
2. Weigert's Hematoxylin Solution A -----125 ml
3. Weigert's Hematoxylin Solution B -----125 ml
4. Aniline Blue Solution -----250 ml
5. Biebrich Scarlet-Acid Fuchsin Solution -----250 ml
6. Phosphomolybdic-Phosphotungstic Acid Solution -----250 ml
7. 1% Acetic Acid Solution -----250 ml

Storage

Room temperature.

Protocol

For formalin-fixed, paraffin-embedded (FFPE) tissue sections:

1. Deparaffinize in Xylene I for 6 minutes and II for 6 minutes.
2. Rehydrate.
 - a. Ethanol 100% (2 minutes)
 - b. Ethanol 100% (2 minutes)
 - c. Ethanol 95% (2 minutes)
 - d. Ethanol 95% (2 minutes)
 - e. Ethanol 70% (2 minutes)
3. Rinse in distilled water (5 minutes).
4. Mordant in Bouin's Solution, 60°C for 1 hour.
5. Wash in running tap water for 5 minutes to remove the picric acid.
6. Prepare Weigert's working hematoxylin: mix Weigert's Hematoxylin Solution A and B at 1:1 ratio.
7. Weigert's working hematoxylin for 10 minutes.
Note: Discard after use.
8. Blue in running tap water for 5 minutes, rinse once for 1 min in distilled water.
9. Biebrich Scarlet-Acid Fuchsin Solution for 5 minutes.
Note: Solution may be used twice only, then discarded.
10. Rinse once for 1 min in distilled water.
11. Phosphotungstic/phosphomolybdic acid solution for 10 minutes, discard solution.
12. Transfer directly into Aniline Blue Solution for 10 minutes.
13. Rinse once for 1 min in distilled water.
14. Acetic Acid Solution for 1 minute, discard solution.
15. Wash slides, 2min x 2 in dH₂O.
16. Dehydrate with 2 changes of 95% Ethanol and 2 changes of 100% Ethanol (2 minute per change).

17. Clear with 3 changes of xylene (5 minutes per change) and coverslip with Permount or other suitable organic mounting medium.

For Frozen Sections:

1. Fix frozen sections in 10% formalin (not Zinc) for 30 minutes.
2. Rinses in distilled water 3×3min.
3. Mordant in Bouin's Solution, 60°C for 1 hour.
4. Wash in running tap water for 5 minutes to remove the picric acid.
5. Weigert's working hematoxylin for 7 minutes.
Note: Discard after use.
6. Blue in running tap water for 5 minutes, rinse in distilled water.
7. Biebrich Scarlet-Acid Fuchsin Solution for 5 minutes.
Note: Solution may be used twice only, then discarded.
8. Rinse in distilled water.
9. Phosphotungstic/phosphomolybdic Acid Solution for 10 minutes, discard solution.
10. Transfer directly into Aniline Blue Solution for 5 minutes.
11. Rinse in distilled water.
12. 1% Acetic acid for 1 minute, discard solution.
13. Wash slides, 2min × 2 in dH₂O.
14. Dehydrate with 2 changes of 95% Ethanol and 2 changes of 100% Ethanol (2 minute per change).
15. Clear with 3 changes of xylene (5 minutes per change) and coverslip with Permount or other suitable organic mounting medium.

Expected results

Cytoplasm, keratin, muscle fibers, Erythrocytes -----	red
Nuclei -----	black
Collagen and mucus -----	blue

Note

This product is intended for research purposes only. This product is **not** intended to be used for therapeutic or diagnostic purposes in humans or animals.

Precautions

Handle with care. Avoid contact with eyes, skin and clothing. Do not ingest. Wear gloves.